

Bioactive glass coatings

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Four kinds of gel-derived materials of the CaO–P₂O₅–SiO₂ (S2, II, I, A2) system were obtained in the form of thin coatings on microscope slides. The obtained materials differed from each other in the ratio of the basic components (CaO and SiO₂). The coatings were characterised with regard to the state of the surface as well as to the phase composition of the materials. In order to determine any bioactive properties of the gel-derived coatings *in vitro*, tests in simulated body fluid (SBF) were made and biochemical examinations using cultured human marrow stromal cells (hMSC) were conducted. It was found that surface crystallisation of hydroxyapatite (HAp) indicating the bioactivity of the material occurred in SBF only in the case of A2 coatings, which are characterised by the highest ratio of CaO:SiO₂. Tests with hMSC showed that the A2 biomaterial promotes both the osteogenesis and remodelling of bone (osteoclastogenesis).

Key words: *biomaterials; bioactivity; sol-gel method*

1. Introduction

Bioactive ceramic materials are based on the CaO–P₂O₅–SiO₂ system [1, 2]. They comprise both glasses and glass-ceramic materials with hydroxyapatite and wollastonite as the main crystal phases. They are assumed to satisfy the following requirements:

- biocompatibility,
- suitability for clinical applications,
- ability to form a stable connection with the bone as well as to promote bone regeneration.

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When in contact with body fluid, these materials form layers of hydroxyapatite (HAp) on their surfaces, through which the implant materials form permanent bonds with the bone in a living organism. The ability to form HAp on the surface of biomaterials is usually indicated by results of simulated body fluid (SBF) tests, which allow *in vitro* character of changes on the biomaterial surface to be estimated after contact with SBF [3]; biomaterials are preliminarily defined by this basic bioactive property.

In recent years, great interest has arisen for a new generation of bioactive materials with increased bioactivity, interpreted mainly as the ability to stimulate faster regeneration of natural tissues [4, 5]. In order to obtain such materials, the sol-gel method is used, which enables biomaterials with a high degree of both chemical and biological surface activities to be produced [6]. This method allows the production of biomaterials in the form of powders and granules, dense and porous sinters, thin coatings, on bioinert substrates. So far, however, it has not been fully recognized which material parameters, such as the state of the surface, pore structure, chemical and phase composition, etc. affect the bioactive properties of these biomaterials. The lack of this knowledge does not permit fully controlled production processes to be carried out.

The aim of this study was to obtain bioactive gel-derived coatings with various characteristics (chemical properties, phase composition, surface roughness) and to evaluate their bioactive properties under *in vitro* conditions from the point of view of material parameters.

2. Materials and methods

Biomaterials from the system $\text{CaO-P}_2\text{O}_5\text{-SiO}_2$ have been chosen for the investigations. Their chemical compositions are given in Table 1.

Table 1. The chemical composition of the investigated materials

Chemical composition	Symbol			
	S2	II	I	A2
SiO_2	80	72	64	40
CaO	16	24	30	54
P_2O_5	4	4	6	6
CaO/SiO_2	0.2	0.33	0.47	1.35

Such a choice of chemical compositions allowed materials to be obtained with various molar ratios of the components having considerable influence on the bioactive properties, i.e. CaO and SiO_2 .

To prepare the starting solutions, the following reagents were used: $\text{Si}(\text{OC}_2\text{H}_5)_4$ (TEOS) (Merck), $\text{OP}(\text{OC}_2\text{H}_5)_3$ (Merck), and $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ (POCh). In addition to

distilled water, ethanol was used as a solvent and hydrochloric acid (HCl) as a catalytic agent. The scheme of preparing the starting solutions is given in Fig. 1.

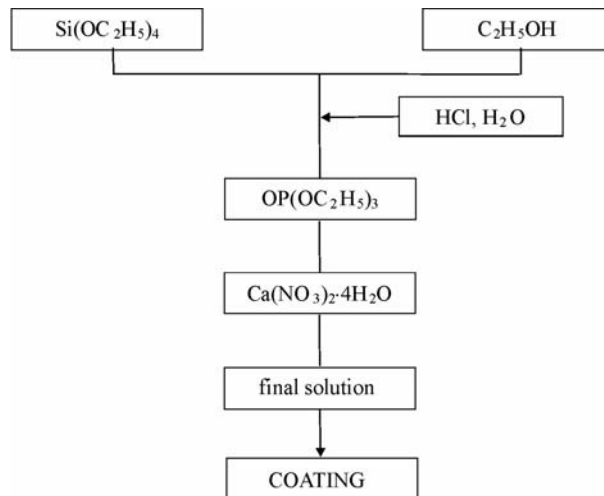


Fig. 1. A scheme for the preparation of coatings

The solutions were used to deposit coatings on bioinert substrates (microscopic glass slides). Thin coatings were deposited by dip coating, using a specially designed apparatus. The glasses with the deposited coatings were dried at ambient temperature and subsequently heated in an electric furnace at the temperature of 450 °C. The coatings obtained were either opaque or transparent. Coatings prepared in this way were subject to the following observations and investigations:

- evaluation of the quality of the coatings by visual and microscopic methods,
- determination of the phase composition of the coating materials by X-ray diffraction analysis (XRD), using a Seifert diffractometer and applying CuK_α radiation;
- determination of layers roughness according to ISO (DIS H287/1), using a profilometer (Hammel Tester T500, Mommelwerke GmbH, Berlin),
- testing bioactivity *in vitro* (the simulated body fluid (SBF) test) comprising:
 - investigation of the solubility of the coating materials in SBF within a period of 1–21 days; to this end, the concentrations of Ca ions in SBF were measured by the complexometric method (applying a solution of disodium versenate in the presence of calcein as an indicator),
 - evaluation of the changes on the surfaces of the coatings after 1–21 days of contact with SBF by means of SEM observations (JEOL 5400, Tokyo, Japan), EDAX analysis (LINK ISIS 300), and XRD diffraction phase analysis (Seifert diffractometer),
 - *in vitro* tests of the A2 material with cultured human marrow stromal cells (hMSC); the aim of this test was to check if the obtained gel-derived material promotes both osteogenesis and bone remodelling (osteoclasogenesis). Three independ-

ent samples of human marrow cells, isolated from the femurs of individuals undergoing total hip replacement, were cultured in a medium (α -MEM + 15% bovine serum FBS). When confluent, the adherent layer of hMSC was re-plated on either the A-2 gel-derived coating or on a tissue-cultured plastic. Cells were harvested for mRNA assays after 7 days with either ascorbate-2-P (Asc), Asc with bone morphogenetic protein BMP2 or dexamethasone (a synthetic glucocorticoid) Dex, or with both BMP2 and Dex. Alkaline phosphatase (ALP) and RANK-L were localised with immunocytochemistry. ALP was treated as a marker of osteogenesis and RANK-L as a marker of the promotion of bone remodelling (osteoclastogenesis) [7–9].

3. Results

3.1. Characteristics of the coatings

The coatings deposited on the microscope slides tightly covered the substrate and were characterized by very good adhesion. Coatings S2, I, and II were translucent, while coatings made from the A2 material were opaque (Fig. 2).

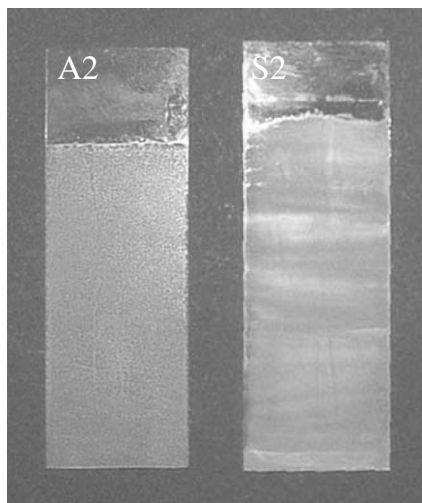


Fig. 2. Thin coatings of the A2 and S2 materials on microscopic glass slides

It can be inferred from SEM observations (Figs. 3, 5) that the quality of the obtained coatings was good. At some places a few cracks, typically in the boundary area, were visible. In the case of the A2 coating, spherical micro areas could be observed, uniformly distributed on the entire surface, evidence of liquation and/or crystallization. The chemical compositions of the coatings were in agreement with the expected compositions of the gel-derived materials (results of EDAX analysis – Figs. 4, 6).

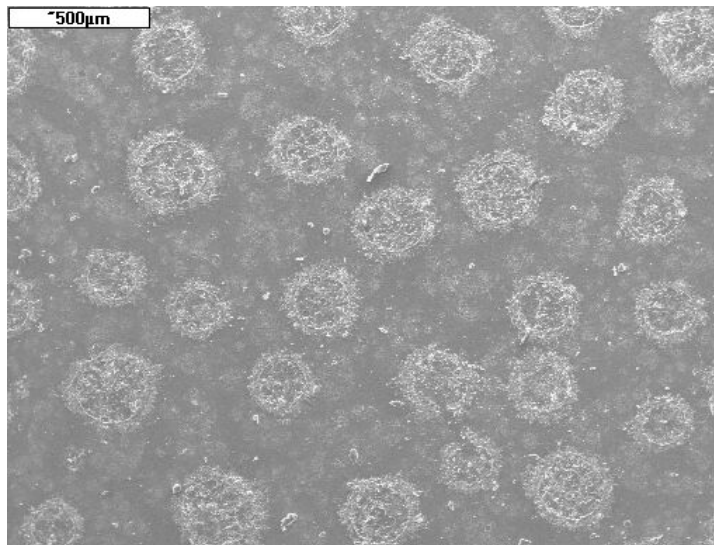


Fig. 3. SEM image of the surface of a gel-derived A2 coating

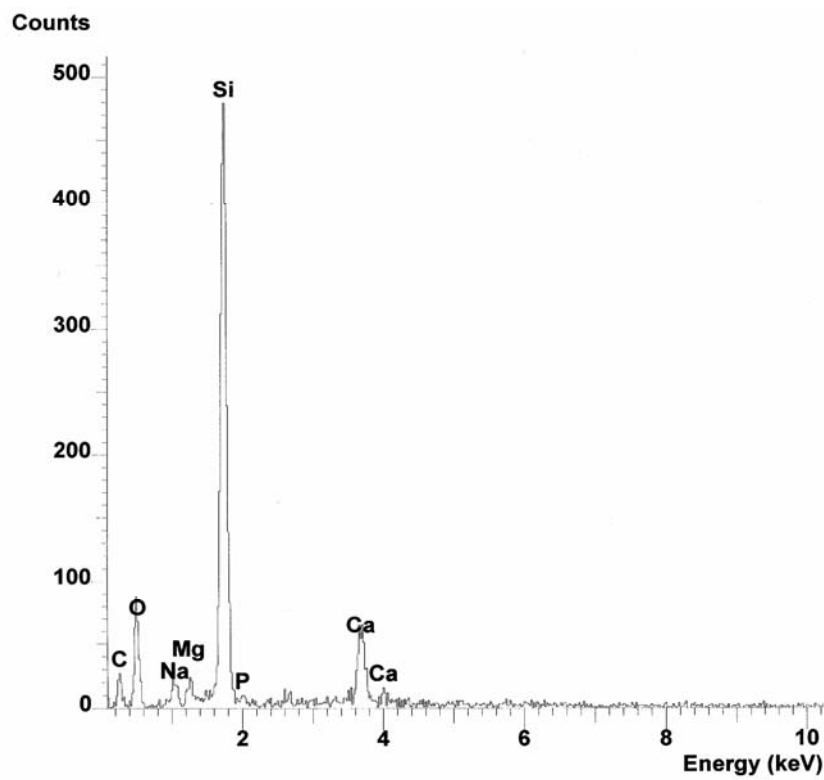


Fig. 4. Results of EDAX analysis for the surface of a gel-derived A2 coating

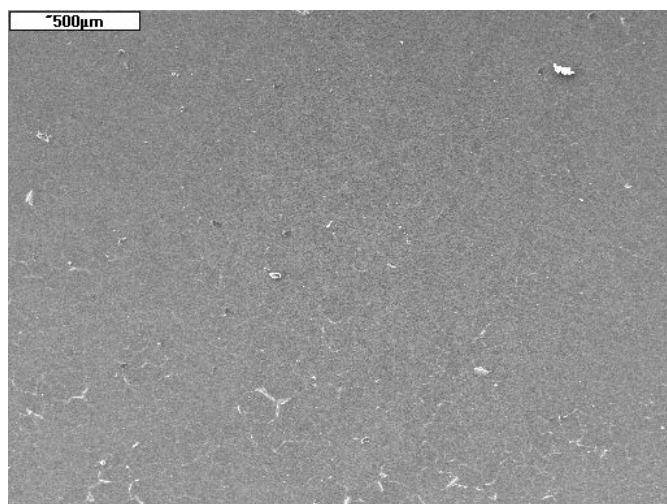


Fig. 5. SEM image of the surface of a gel-derived S2 coating

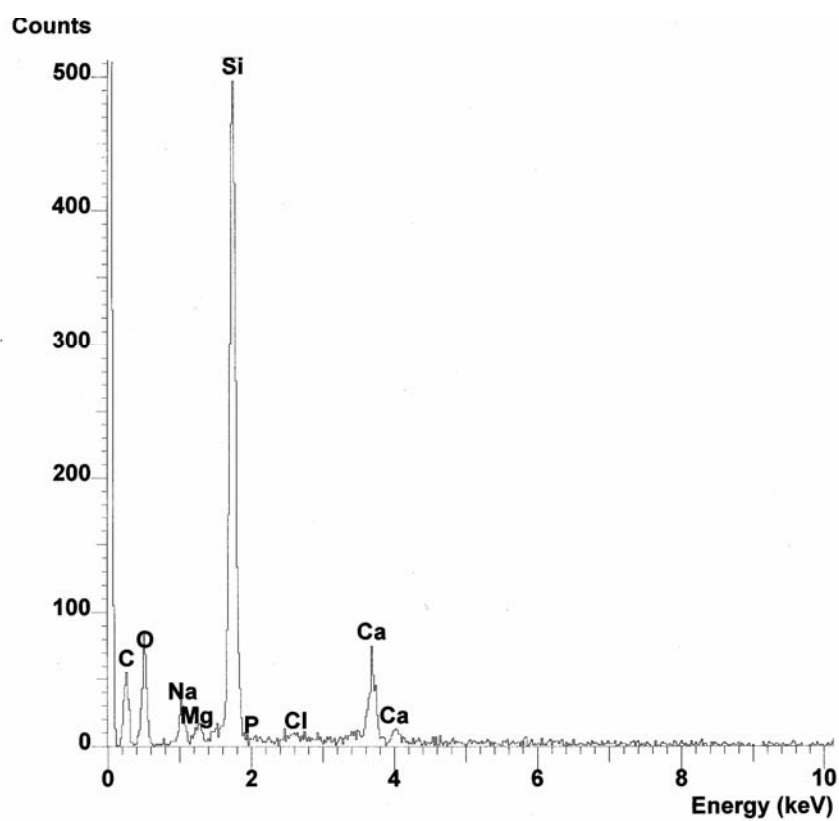


Fig. 6. Results of EDAX analysis for the surface of a gel-derived S2 coating

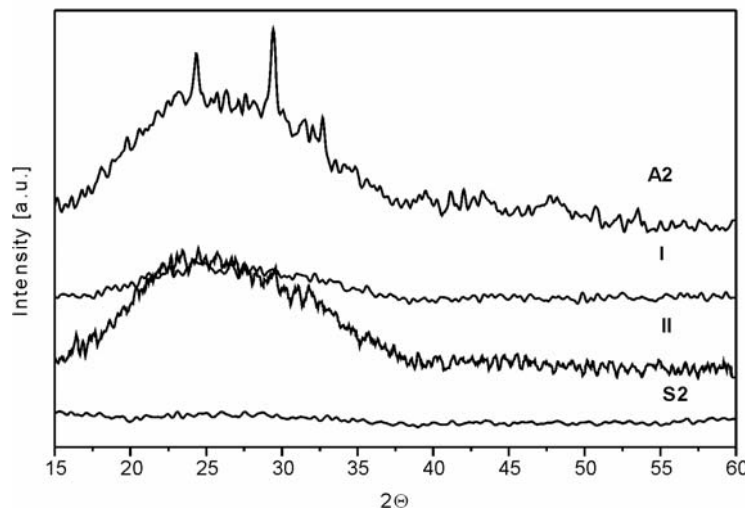


Fig. 7. XRD patterns of gel-derived A2, II, I, and S2 coatings

X-ray diffraction analysis showed (Fig. 7) that the coatings S2, I, and II are completely amorphous, while the coating made of the A2 material contains a crystal phase (phases) with calcium. On account of the low intensity of the reflexes, however, it is difficult to identify the character of these phases (phase).

Table 2. The surface properties of the investigated materials

Coefficient	Gel-derived layer			
	S2	A2	I	II
R_a	0.06	0.96	0.06	0.12
R_z	0.59	10.19	0.80	1.83
R_t	1.72	11.71	1.10	4.17

Three parameters defining the state of the surface coatings (roughness and topography) have been measured (Table 2):

- R_a – the arithmetic mean of the deviation of the filtered roughness profile from the centre line within the measured length;
- R_t – the vertical distance between the maximum and the highest points in the filtered roughness profile within the reference length;
- R_z – the height of ten points (upper level of the absolute values of the five highest peaks and five lowest valleys within the measured length 1m).

From the data given in Table 2, it appears that the A2 coating, in which crystallization occurred, was characterized by the largest roughness. In the case of the amorphous coatings S2 and I, the roughness was considerably smaller, whereas the amor-

phous coating II exhibited a roughness that could be defined as intermediate between that of A2 and the materials S2 and I.

3.2. Test in SBF

Treatment of the gel-derived coatings with simulated body fluid (SBF) induced some changes in the chemical composition of the fluid (Fig. 8) as well as on the surface of the coatings (results of EDAX analysis – Figs. 10, 12).

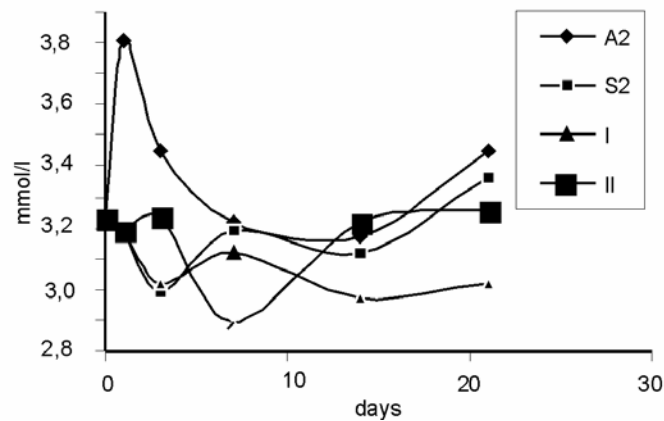


Fig. 8. Ca concentration of glass slides covered with the gel-derived A2, II, I, and S2 materials after various immersion times in SBF

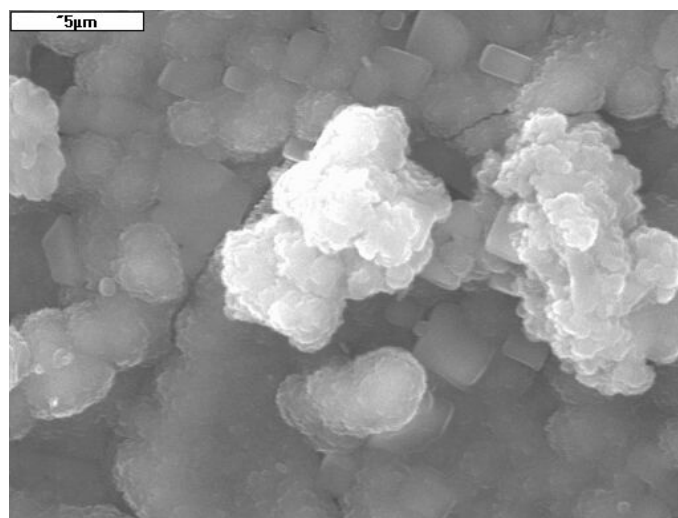


Fig. 9. SEM image of the surface of a gel-derived A2 coating after 7 days contact with SBF

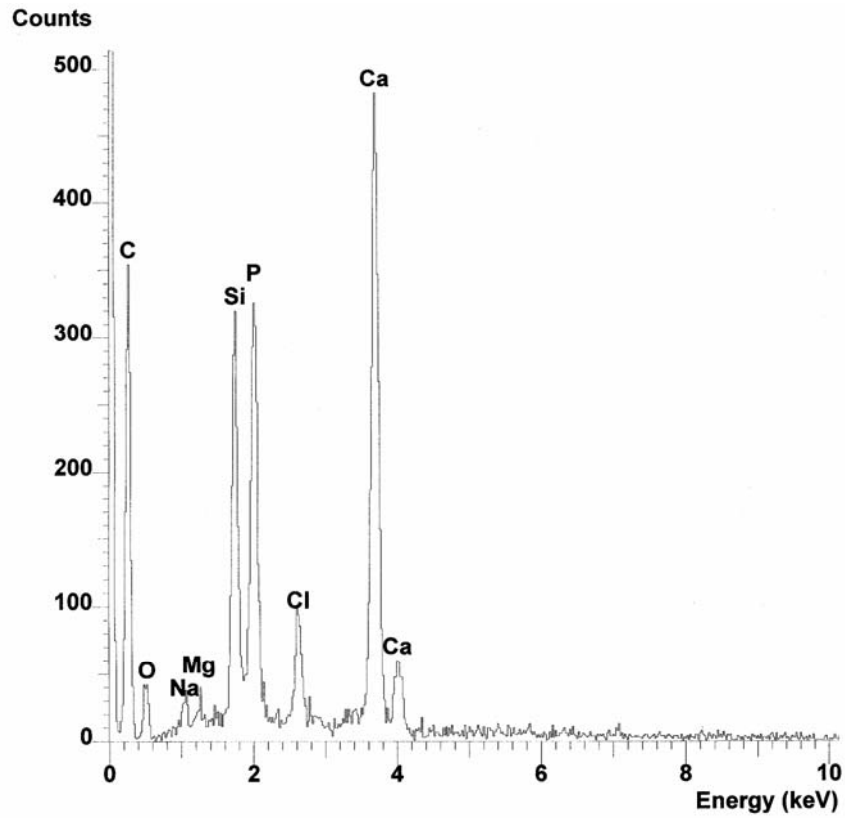


Fig. 10. Results of EDAX analysis for the surface of a gel-derived A2 coating after 7 days contact with SBF

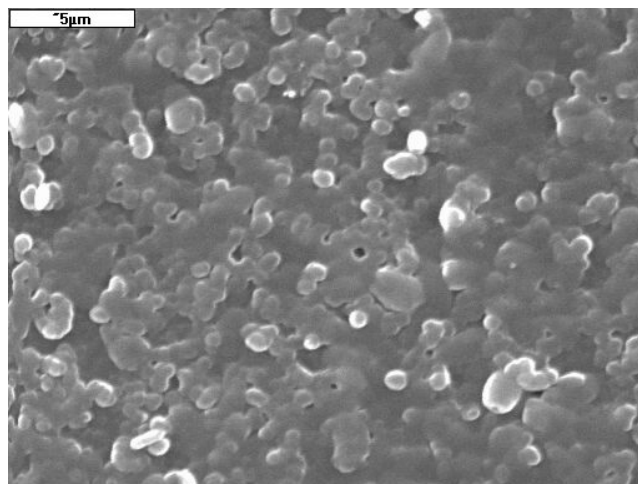


Fig. 11. SEM image of the surface of a gel-derived S2 coating after 7 days contact with SBF

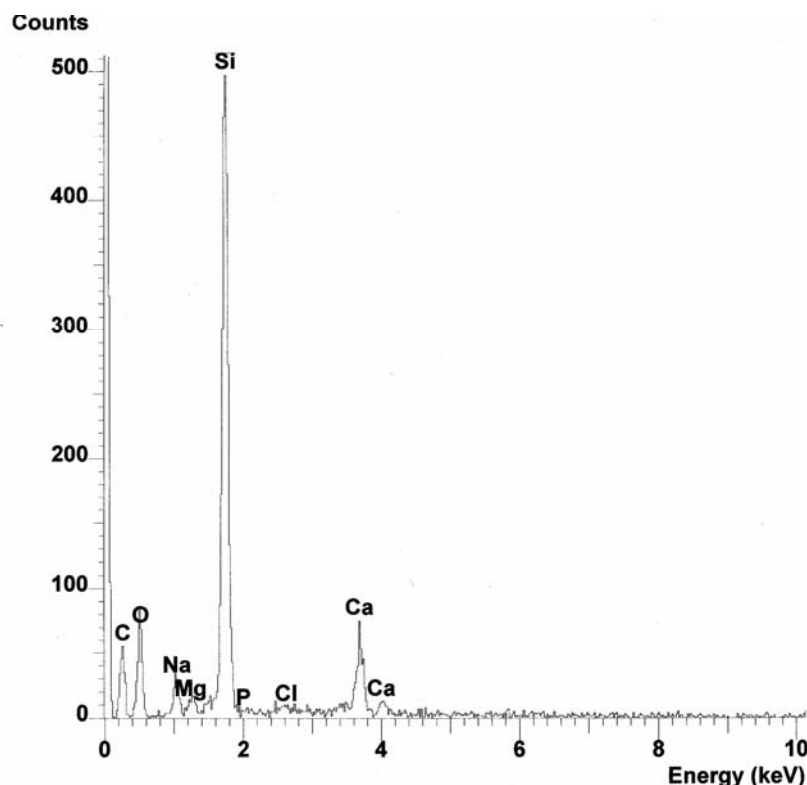


Fig. 12. Results of EDAX analysis for the surface of a gel-derived S2 coating after 7 days contact with SBF

Changes in SBF composition were caused by the solubility of the coating materials. This solubility was evaluated only with respect to the calcium content in SBF. In the case of other components of the coatings, no measurable solubilities have been observed. From among the four examined gel-derived materials, only the A2 coating, for which the $\text{CaO}:\text{SiO}_2$ ratio is the highest, exhibited a considerable loss of calcium already 1 day after contact with SBF. The consequence of this was an increase in the Ca concentration in the SBF. With further contact, the content of Ca in SBF showed a tendency to fall, which might be connected to the surface crystallization of hydroxyapatite. In the case of other coatings, the Ca concentration in SBF, independently of the duration of contact with the biomaterial, was close to the starting concentration.

Changes in chemical composition and SEM images (resulting from contact with SBF) were observed only in the case of the A2 material (Figs. 9, 10). From EDAX analysis (Fig. 10) it appears that already after 7 days after immersion, the concentration of calcium and phosphorus in the surface layer of the material increased. This is an indication of the surface crystallization of calcium phosphates. With prolonged contact time with SBF (up to 21 days), the content of Ca and P on the surface increased, while the content of silicon decreased.

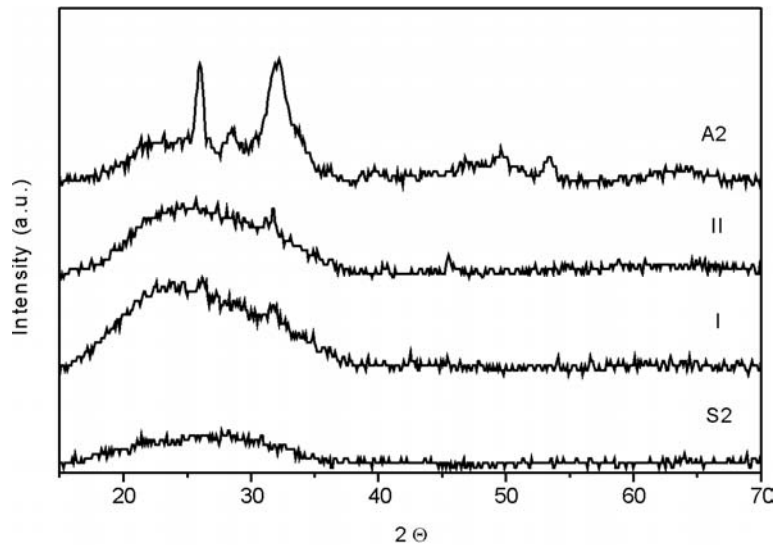


Fig. 13. XRD patterns of gel-derived A2, II, I, and S2 coatings after 7 days contact with SBF

Changes in the chemical composition were accompanied by a distinct change in the morphology of the layer surface. The surface became covered with spherical forms (Fig. 9), which after 21 days completely covered the primary layer. XRD investigations (Fig. 13) showed that the spherical forms appearing on the surface were composed of hydroxyapatite. The described phenomenon was observed for layers characterized by a lower ratio of CaO to SiO₂ (Figs. 11–13). Since the results of the tests with SBF show that only the A2 material is able to produce hydroxyapatite on its surface, it was chosen for *in vitro* tests with human marrow cells.

3.3. In vitro test with human marrow stromal cells

Cells cultured directly on the A2 surface showed a stimulation pattern for alkaline phosphates (ALP) similar to that of cells cultured on plastic, though the levels on A2 were slightly lower (Fig. 14a). The A2 glass, however, had a far more profound effect on the expression of osteopontin and RANK-L (Fig. 14b). MSC cultured on a A2 surface had osteopontin and RANK-L mRNA levels 5–10 times higher than those of cells on plastic, and addition of BPM caused a further increase in the mRNA levels.

4. Discussion

The sol-gel process enabled coatings with various chemical compositions to be obtained in a relatively simple way on glass plates. The phase composition of the layers depended on the molar ratio CaO:SiO₂. At the highest value of this ratio (ca. 12), the

liquation and crystallization of the phase enriched with calcium probably took place. The effect of crystallization was the deterioration of surface smoothness, resulting in an increase of the parameters R_a , R_z , and R_t .

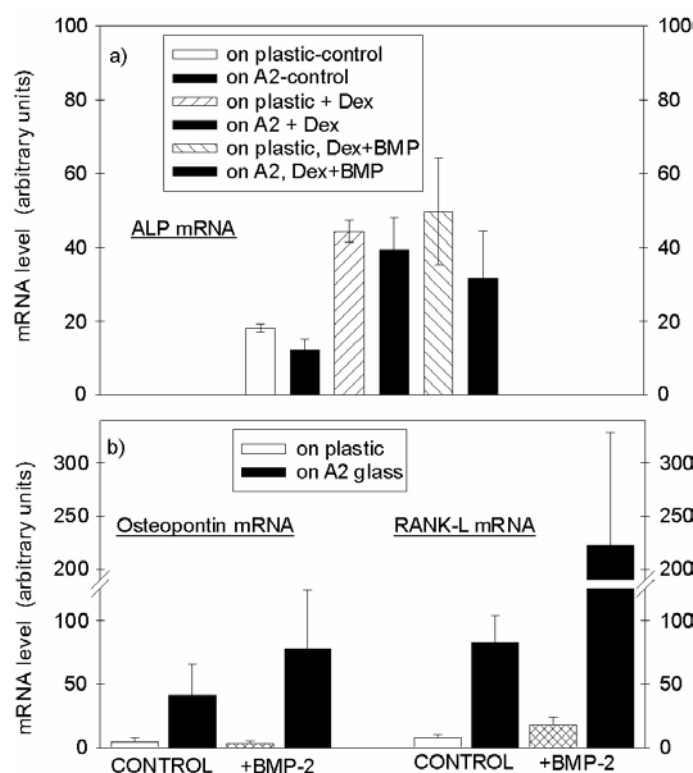


Fig. 14. Expression of mRNA in hMSC, cultured for 7 days on a tissue culture plastic (plastic) and on A2 gel-derived coating (A2 glass, black bars). Cells were cultured with a media containing 100 μ g/ml ascorbate (control), a control media with 100 nM Dex, a control media with 100 μ g/ml BMP-2, or with Dex + BMP. Comparison of ALP mRNA in cells on plastic or A2 glass with various inducers (A). The effect of A2 glass, with and without BMP treatment, on the expression of mRNA for osteopontin and RANK-L (B)

When considering the chemical composition of the examined layers from the point of view of its influence on the bioactive properties of the materials, it should be expected that increasing the SiO_2 concentration would favour HAp crystallization due to an increase of the number of active nucleation centres in the form of Si-OH groups [10, 11]. On the other hand, HAp crystallization can occur at an appropriate concentration of calcium in SBF, exceeding the solubility HAp. Thus, the increased concentration of calcium in the material should be a factor promoting HAp crystallization in this case.

As a result of the test in SBF, it has been found that within the investigated period of 21 days HAp was formed due to contact with SBF only on the surface of the A2

material, which is characterized by the highest CaO:SiO₂ ratio. At the same time, only in A2 layers a considerable solubility of Ca in SBF was observed. Thus, it can be assumed that an appropriate level of calcium solubility, determined by the value of the CaO:SiO₂ ratio in CaO–P₂O₅–SiO₂ gel-derived coatings, is the factor that determines the ability of HAp to form on the surface. Accordingly, it can be expected that coatings with appropriately high CaO:SiO₂ ratios can be bioactive. This statement, however, needs to be verified in *in vitro* conditions.

At the same time, our *in vitro* study identifies the bioactive gel-derived A2 material to potentially support the growth and osteoblast differentiation of human bone marrow stromal cells. Surfaces coated with the A2 sol-gel glass were found to permit the adherence, proliferation, and differentiation of hMSC at levels comparable to those seen for tissue culture plastics. Furthermore, the A2 glass induces the expression of both osteopontin, which promotes the migration of osteoblast precursors, and RANK-L, which induces the differentiation of osteoclast precursors. Our data indicate that those MSC that exhibit increased ALP levels are the same cells that show elevated RANK-L levels. There may be, however, an inverse relationship between the levels of ALP and RANK-L. The ability to promote both osteogenesis and osteoclastogenesis may facilitate not only new bone formation, but also the subsequent remodelling of the new bone. Since hMSC are believed to be stem cells that play a key role in providing new osteoblasts for bone repair and remodelling, A2 gel-derived glass is significantly promising as a coating for orthopaedic and dental implants.

5. Conclusions

The bioactive properties of CaO–P₂O₅–SiO₂ gel-derived coatings, determined on the basis of HAp surface crystallization capability caused by contact with simulated body fluid (SBF), are determined for different CaO:SiO₂ ratios. The increase of CaO content at the expense of SiO₂ leads to a higher solubility of Ca in SBF and promotes the surface crystallization of HAp.

The material for which the surface crystallization of HAp occurs as a result of contact with SBF, promotes both osteogenesis and osteoclastogenesis, which may facilitate the formation and remodelling of the newly formed bone.

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